Oral Biofilm Architecture on Natural Teeth

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Introduction

Oral microbial biofilms are three-dimensional structured bacterial communities [1] attached to a solid surface like the enamel of the teeth, the surface of the root or dental implants [2] and are embedded in an exo-polysaccharide matrix [3]. Oral biofilms are exemplary and served as a model system for bacterial adhesion [4,5] and antibiotic resistance [6].

The appreciation of the complex nature of oral biofilms was highlighted decades ago by the work of Listgarten and co-workers who described the architecture of biofilms by light and electron microscopy on epoxy resin crowns and extracted teeth [7,8]. Supragingivally, on the enamel, they observed the formation of columnar micro-colonies with their long axis perpendicular to the crown surface. Gram-positive cocci dominated these columns and were embedded in an exo-polysaccharide matrix [3]. Oral biofilms provide a clear vision on biofilm architecture and the spatial distribution of predominant species.

Abstract

Periodontitis and caries are infectious diseases of the oral cavity in which oral biofilms play a causative role. Moreover, oral biofilms are widely studied as model systems for bacterial adhesion, biofilm development, and biofilm resistance to antibiotics, due to their widespread presence and accessibility. Despite descriptions of initial plaque formation on the tooth surface, studies on mature plaque and plaque structure below the gum are limited to landmark studies from the 1970s, without appreciating the breadth of microbial diversity in the plaque. We used fluorescent in situ hybridization to localize in vivo the most abundant species from different phyla and species associated with periodontitis on seven embedded teeth obtained from four different subjects. The data showed convincingly the dominance of Actinomyces sp., Tannerella forsythia, Fusobacterium nucleatum, Spirochaetes, and Synergistetes in subgingival plaque. The latter proved to be new with a possibly important role in host-pathogen interaction due to its localization in close proximity to immune cells. The present study identified for the first time in vivo that Lactobacillus sp. are the central cells of bacterial aggregates in subgingival plaque, and that Streptococcus sp. and the yeast Candida albicans form corncob structures in supragingival plaque. Finally, periodontal pathogens colonize already formed biofilms and form microcolonies therein. These in vivo observations on oral biofilms provide a clear vision on biofilm architecture and the spatial distribution of predominant species.


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and two phyla without cultivivable members; OP11 and TM7, which is summarized in Figure 1. Little is known about the spatial distribution of these taxa in oral biofilms. The aim of the present study therefore was to reveal the in vivo architecture of supra and subgingival plaque with a panel of 16S or 18S rRNA targeted FISH-probes covering the most important groups of oral microorganisms, and to provide an essential step from oral microbial diversity to oral biofilm function.

**Results**

From 10 examined teeth, seven showed a positive signal after hybridization with fluorescently labeled probes. Macroscopic analysis of Gram-stained sections revealed the localization of the plaque in relation to the cemento-enamel junction and gingival tissue. A phylogenetic tree based on approximately 1500 nearly complete (>1300 bp) sequences was constructed. Sequences were derived from molecular studies of oral microbial diversity and a manual search through the SILVA database [12]. The coverage of the applied probes is represented in Figure 1. It shows that a representative part of the oral microbial diversity is covered.

**Subgingival Biofilm Architecture**

The localization of the most abundant subgingival bacteria is summarized in Figure 2. Panel A shows a typical subgingival biofilm with increasing fluorescent intensity from the tooth side towards the epithelium side. Based on differences in bacterial morphologies and fluorescence intensities, four different layers were distinguished. The first layer of the biofilm is composed of cells displaying little fluorescence relative to cells in the top of the biofilm. Of all the probes tested, only *Actinomyces* sp. gave a positive signal in this layer. The intermediate layer is composed of many spindle-shaped cells of which *F. nucleatum*, *T. forsythia* and possibly other *Tannerella* sp. positive with probe Tfor127 are visible as the red/yellow band of bacteria in panels E and F of Figure 2. The top layer of the biofilm and part of the intermediate layer contain mainly bacteria belonging to the Cytophaga-Flavobacterium-Bacteroides cluster (CFB-cluster) as detected with probe CFB935 and shown in panel D. CFB935 positive cells are filamentous, rod-shaped or even coccoid. Samples double stained with CFB935 and *Tannerella*-specific probes showed that most filamentous bacteria are *Tannerella* sp., while many of the rod-shaped bacteria are *Prevotella* sp. and *Bacteroides* species as detected with the group-specific probes PREV392 and BAC303, respectively. Besides the presence of bacteria from the CFB-cluster, large cigar-like bacteria were in the top layer. These cells belong to the Synergistes group A of bacteria and form a ‘palisade’-like lining. They were in close contact to eukaryotic cells resembling polymorphnuclear leukocytes (PMN’s) according to the presence of polymorph nuclei (panel C). Outside the biofilm, a fourth layer without clear organization was observed. *Spirochaetes* were primary localized in the fourth layer where they are the most dominant species. Bacterial aggregates, called rough and fine test-tube brushes were detected between the *Spirochaetes* (Panel B).

**Supragingival Biofilm Architecture**

Supragingival biofilms are more heterogeneous in architecture compared to subgingival biofilms. In general, two different layers could be observed. The basal layer adheres to the tooth surface and four different biofilm types were observed (Figure 3). First, a biofilm composed of only rod shaped *Actinomyces* cells perpendicularly orientated to the tooth surface (panel D). The second type is a mixture of *Actinomyces* sp. and chains of cocci, not identified as streptococci, perpendicularly orientated to the tooth surface (panel E). The third type shows a biofilm with filamentous bacteria, streptococci and yeasts, where streptococci form a distinct colony around yeast cells (panel F). The fourth type is a biofilm composed of mainly streptococci growing in close proximity to *Lactobacillus* sp. that are orientated perpendicularly to the tooth surface (panel G).

The second layer can be found on top of any biofilm type of the basal layer. *Streptococcus* sp. can be present as heterogeneously scattered cells through the second layer of the biofilm without any apparent organization (panel A3), or they can be aligned on top of the second biofilm layer as a thin coat (panel A1). In addition, they colonize cracks in the biofilm (panel A2). Also, there is a heterogeneous scattering of bacterial cells belonging to the CFB-cluster (panel B). Finally, *Lactobacillus* sp. that are orientated away from the tooth surface are surrounded by cells with different morphologies (panel C).
Subgingival Localization of Periodontal Pathogens

The localization of presumptive periodontal pathogens in the subgingival biofilm is shown in Figure 4. Most of the periodontal pathogens belong to the gram-negative group of bacteria united in the CFB-cluster like *P. gingivalis*, *P. intermedia*, *P. endodontalis* or *P. nigrescens*. Most CFB-cluster cells are evenly distributed in the top and intermediate layer of the biofilm. *Prevotella* sp. however, colonize the biofilm in micro-colonies (panel A) which sometimes are located on top of the biofilm, but, as is shown for *P. intermedia*, also reside within the top layer (panel E). *Porphyromonas gingivalis* and *Porphyromonas endodontalis* appear mainly as micro-colonies within the top layer (panel C and D). *Parvimonas micra*, an example of a gram-positive species that is associated with periodontitis, is also found in micro-colonies in the top layer (panel B). Apparently, the microorganisms considered pathogens are mostly present in micro-colonies in the top layer and in the fourth layer of the subgingival biofilm or can be part of bacterial aggregates.

Bacterial Aggregates or Structures in Dental Plaque

Aggregates of microorganisms have been detected in both sub and supragingival plaque (Figure 5). In line with previous reports, different aggregate morphologies were observed in the fourth layer of subgingival plaque. Filaments from the CFB-cluster, morphologically like *T. forsythia*, and *F. nucleatum* are arranged perpendicularly around lactobacilli, forming fine test-tube brushes (panel A–C). There have also been observations of test-tube brushes composed of a complex mixture of cells with *T. forsythia*, *Campylobacter* sp., *P. micra*, *Fusobacteria* and Synergistetes group A, among others. *Synergistetes* cells may also form aggregates exclusively with themselves (panel D and E). In supragingival

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**Figure 2. Localization of the most abundant species in subgingival biofilms.** (A) Overview of the subgingival biofilm with *Actinomyces* sp. (green bacteria), bacteria (red) and eukaryotic cells (large green cells on top). (B) *Spirochaetes* (yellow) outside the biofilm. (C) Detail of *Synergistetes* (yellow) in the top layer in close proximity to eukaryotic cells (green). (D) CFB-cluster (yellow) in the top and intermediate layer. (E) *F. nucleatum* in the intermediate layer. (F) *Tannerella* sp. (yellow) in the intermediate layer. Each panel is double-stained with probe EUB338 labeled with FITC or Cy3. The yellow color results from the simultaneous staining with FITC and Cy3 labeled probes. Bars are 10 µm.

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Discussion

The aim of the present study was to unravel tooth attached biofilm architecture. For the first time, bacteria in these biofilms are identified and localized in vivo using FISH. The results present new insights into the architecture of tooth-attached biofilms and visualize the interaction of the biofilm with the human immune system. FISH offers the opportunity to obtain positional information of bacteria in intact biofilms, its application overcomes the limits of culturability and can relatively easily be extended to new identified species and phylotypes.

Synergistetes sp. for example, only recently gained attention [13] but may account for 3–11% of the bacterial population [14]. In addition, they form a palisade-like lining along the outer length of the biofilm and were in direct contact with host immune cells suggesting an important role in host-biofilm interactions.

In our current experimental design, group-level probes and species-specific probes were applied to efficiently identify cells that might be of interest due to their location in the biofilm. Panels A, B, C, E, F are double stained with probe EUB338 labeled with FITC or Cy3. Bars are 10 μm.

doi:10.1371/journal.pone.0009321.g003

Figure 3. Localization of the most abundant species in supragingival biofilms. Streptococcus sp. (yellow) form a thin band on top of the biofilm (A1), almost engulfing in the biofilm (A2) or present as small cells scattered through the top layer of the biofilm (A3). (B) Cells from the CFB-cluster of bacteria in the top layer of the biofilm, without defined structure. (C) Lactobacillus sp. (red) forming long strings through the top layer. (D) Actinomyces sp. (yellow) plaque attached to the tooth. (E) Actinomyces sp. (green) and cocci forming initial plaque. (F) Multispecies initial plaque composed of Streptococcus sp. (yellow), yeast cells (green) and bacteria unidentified (red). (G) Streptococcus sp. (green) and Lactobacillus sp. (red) forming initial plaque. Black holes might be channels through the biofilm. Panels A, B, C, E, F are double stained with probe EUB338 labeled with FITC or Cy3. Bars are 10 μm.
without quantitative measurements or dynamic time lapse observations. Combining multiple observations of supra- and subgingival biofilms reflects to some degree the dynamics of biofilm formation, which leads us to the following view of plaque formation.

Initial plaque formation starts with the deposition of a salivary pellicle on the tooth surface. Planktonic cells or aggregates of cells adhere to this pellicle via specialized adhesins on the bacterial cell surface that recognize pellicle proteins [16] and by non-specific physico-chemical interactions [5]. These phenomena may result in a scattered pattern of bacterial deposits [9,17] composed of initial colonizers like Actinomyces sp., Streptococcus sp., Lactobacillus sp. and Candida sp. [18–20] and is reflected in the different biofilm types of the first layer of supragingival plaque (Figure 3). Maturation of the biofilm proceeds via co-aggregation of planktonic bacteria to the already adhered biofilm [21] and bacterial growth, as has been shown for Streptococcus sanguinis [22]. The second layer may be the result of both processes. The presence of either Streptococcus sp. or bacteria from the CFB-cluster in the second layer of Figure 3 may reflect a crucial transition in supragingival plaque from a predominantly gram-positive saccharolytic plaque to a gram-negative proteolytic plaque that might be the result of the availability of nutrients e.g. dietary sugars or proteins from saliva and crevicular fluids. It was noticed that after three weeks, undisturbed supragingival plaque morphologically resembled subgingival plaque [7]. In our observations of subgingival plaque, the fluorescence intensity of the bacterial cells stained with the eubacterial probe increased from the tooth side towards the epithelium. This reflects differences in physiological activity of the cells [23]. In the basal layer of Figure 2, only Actinomyces sp. showed positive of all the probes tested. The unidentified cells may belong to new species for which no probes have been developed. Another explanation might be that the basal layer constitutes previous stages of the biofilm that have become secluded from nutrients and contain dead or physiologically inactive cells with lower fluorescence intensity as has been shown in vitro [24,25]. Of the initial colonizers, only Actinomyces sp. might survive, maybe due to their capacity to store intracellular glycogen [26] or their capacity to scavenge on biofilm material like extracellular polymeric sub-

Figure 4. Localization of species associated with periodontitis. (A) Overview of the subgingival biofilm with CFB-cluster species (red) and Prevotella sp. (yellow). Since Prevotella sp. are part of the CFB-cluster of bacteria, cells appear in yellow. (B) Top of the biofilm with a micro-colony of P. micra (yellow). (C) Micro-colonies of P. gingivalis (yellow) in the top layer. (D) Micro-colonies of P. endodontalis (yellow) in the top layer. (E) Micro-colonies of P. intermedia in the top layer. Panels B, C, D and E are double stained with probe EUB338 labeled with FITC or Cy3. Bars are 10 μm. doi:10.1371/journal.pone.0009321.g004
stances and on compounds from dead bacterial cells. These are the first in vivo observations of graduated differences in the physiological activity of cells within the biofilm, and support the idea that bacterial growth is an important determent of oral biofilm development [27].

In the intermediate layer, T. forsythia may benefit from its close proximity to dead cells in the basal layer. These may serve as a source of exogenous N-acetyl-muramic acid, a bacterial cell wall sugar on which T. forsythia is dependent [28]. In the presence of F. nucleatum, T. forsythia synergistically forms robust biofilms via cell-cell contacts in vitro [29] as is reflected in their prominent and abundant co-localization of both species along the entire length of the biofilm. The presence of F. nucleatum in the intermediate layer, as proposed by Kolenbrander and London [15], is confirmed for the first time in vivo in the present study. These structural observations on the dominance of Actinomyces sp., Fusobacteria and T. forsythia are supported by dot-blot analysis of subgingival plaque [30]. In contrast, P. gingivalis, P. intermedia, P. endodontalis and P. micro are mainly located in the top layer of the biofilm in micro-colonies [31,32]. The presence of micro-colonies proposes a distinction between species that are structurally present, probably forming the framework of the biofilm, and transient species that can colonize the already established biofilm forming micro-colonies.

Summarizing, the present study on oral biofilms links early studies on biofilm structure and recent molecular insights in oral bacterial diversity. This resulted in important new observations on oral biofilms, in architecture and in dynamics. First, the species...
Table 1. Oligonucleotide probe sequences, their targets and the hybridization conditions used in this study.

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<th>Target</th>
<th>Probename</th>
<th>Label</th>
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<th>T\textsubscript{m} (°C)</th>
<th>Pretreatment</th>
<th>Formamide (%)</th>
<th>Hyb. Temp. (°C)</th>
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<td>lysis buffer</td>
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<td>60 min 37°C</td>
<td>labmix</td>
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<td>46</td>
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<td>40–50</td>
<td>46</td>
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<td>46</td>
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New probes developed in the present study have been designed with the ARB software package [46] and tested against a panel of reference strains for specificity (ACT218, Aa1829, LAB759, Sel1469 and Fna1254) or tested on subgingival plaque samples at increasing formamide concentrations to define assay conditions for maximum stringency and optimal specificity (TrepG1, Pendo740 and SynA1409). Probe LAB759 showed cross-reactivity with Eikenella corrodens and not identified cocci. In the present study, only rods gave a positive signal with probe LAB759.

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that form test-tube brushes and cornsobs are identified for the first time in vivo. Second, the localization of T. forsythia in the intermediate layer of oral biofilms should be incorporated in the biofilm model, as well as a fourth layer of unattached plaque consisting of mainly Sporohales. Third, the observation of bacteria that are either structural members of the subgingival biofilm, e.g. Actinomyces, Fusobacteria, Tannerella sp., or species that colonize an already formed biofilm, e.g. P. intermedia, P. gingivalis and P. micro. Fourth, the biofilm model based on co-aggregation should include bacterial growth and appreciate the dynamics of biofilm maturation. Finally, the finding of a palisade lining of Syngnogtis sp. in the close proximity to host defence cells suggests a major role in host biofilm interactions. These results provide an oral biofilm model and show that in vivo observations on biofilm architecture are an essential link between molecular diversity and bacterial function in relation to oral diseases.

Materials and Methods

Ethics Statement

All protocols were approved by the Medical Ethical board of the University Medical Center Groningen. Extracted teeth were collected as anonymous by-products of regular treatment. As such, the Medical Ethical board stated that the performed research was not conducted under the regulations of the Act on Medical Research Involving Human Subjects (METC 2009.305).

A written informed consent was therefore not compulsory. Nevertheless, patients were informed about the research purposes and gave verbal informed consent, which was not recorded to keep the procedure anonymously.

Sample Collection and Handling

Ten teeth from four patients were used in this study. The patients were referred to the Dept. of Oral Surgery and Periodontology for extraction of their remaining teeth and the fabrication of complete dentures. Teeth were diagnosed with advanced generalized periodontitis based on pocket depth recordings of >6 mm and x-rays indicating more than 30% bone loss. Subjects had not taken antibiotics within the last three months and did not suffer from systemic diseases. An experienced dentist carefully extracted the teeth without the use of elevators, not to disturb the subgingival plaque. Immediately after extraction, the teeth were placed in 3% (wt/vol) paraformaldehyde (PFA) in phosphate-buffered saline (PBS) (8 g of NaCl, 0.2 g of KCl, 1.44 g of Na2HPO4 and 0.24 g of KH2PO4 per liter; pH 7.2) and fixed at 4°C until further use or processed immediately.

Sample Processing

Fixed and dehydrated teeth were carefully embedded in Technovit 8100 (Heraeus Kutzler GmbH) at 4°C. The embedded tissue was either cut transversal or a combination of longitudinal and transversal (50/50) in cross-sections of 1 mm with a water-cooled rotary saw. The embedded cross-sections were decalcified with a 17% ethylene-diamine-tetraacetic acid decalcifying solution (pH 7.0), which was renewed regularly during the course of decalcification, varying in duration from 16 to 22 days depending on the size of the specimens. Regular x-ray analysis confirmed completion of decalcification. Decalcified cross-sections were re-embedded in Technovit 8100 and stored at 4°C. Sections of two micron were obtained with a Tungsten carbide knife in a rotary microtome (Reichert-Jung) and stretched on water. Stretched sections were mounted to polysine precoated glass slides (Thermo Scientific) for FISH analysis.

FISH Analysis

Oligonucleotide probes, labeled at the 5’- and 3’- end with fluorescein (FITC) or at the 5’- end with Cy3 were purchased from Eurogentec (Eurogentec, Maastricht, the Netherlands). A set of 29 FISH probes, specific at the domain or group level were used together with species-specific probes (Table 1). The target bacteria of probe LAB759 needed pre-treatment with Labmix (25 mM Tris-HCl pH 7.5, 10 mM EDTA, 585 mM sucrose, 5 mM CaCl2, 0.3 mg/ml sodiumtaurocholate, 0.1 mg/ml lipase and 2 mg/ml lysisome) for 1 h at 37°C. To enable probe penetration, other gram-positive targets needed lysozyme pre-treatment for 15 min at room temperature with lysozyme buffer (2 mg/ml lysozyme, 100 mM Tris-HCl, pH 8.0) as indicated in Table 1. Standard FISH procedures were followed with a hybridization time of three hours and formamide concentration and hybridization temperature (46°C or 50°C) according to the references (Table 1), with the aim of achieving optimal stringency and specificity. The biofilms were examined using a Leica DM RXA microscope (Leica Mikroskopie). Filters were set to 500–540 nm for FITC and 570–630 nm for Cy3. Images were obtained using 63x (numeric aperture 1.0) oil immersion objectives. Color micrographs were taken with a digital Canon EOS400 Camera, transferred to an HP personal computer and processed using Photoshop 6.0 (Adobe) without any qualitative changes to the raw images.

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Author Contributions

Conceived and designed the experiments: VZ HJH. Performed the experiments: VZ BvL TT RG. Analyzed the data: VZ RG HJH. Contributed reagents/materials/analysis tools: VZ BvL HJH. Wrote the paper: VZ JED FA RG HJH.

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